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Research Article

# Antiulcer Activity of the Root Bark of *Oroxylum indicum*. Against Experimental Gastric Ulcers

Maitreyi Khandhar, Mamta Shah, Devdas Santani & Sunita Jain

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## Abstract

The current study was undertaken to investigate the effect of the root bark of *Oroxylum indicum* Vent. (Bignoniaceae) against experimental gastric ulcers. The 50% alcohol extract of root bark of *Oroxylum indicum*. and its different fractions, viz., petroleum ether, chloroform, ethyl acetate and n.-butanol, were studied (p.o.) against ethanol-induced gastric mucosal damage. The alcohol extract (300 mg/kg) and its different fractions (100 and 300 mg/kg) showed significant reduction in gastric ulceration. Out of all these fractions, the petroleum ether (96%) and n.-butanol (99%) fractions showed maximum inhibition of gastric lesions against ethanol-induced gastric mucosal damage. The results were comparable with omeprazole (reference standard). In the ethanol-induced gastric ulcer model, treatment with both the active fractions and omeprazole

lipid peroxidation that was measured in terms of malondialdehyde (MDA), along with significant rise in the superoxide dismutase (SOD), catalase (CAT), and reduced glutathione levels (GSH), when compared with the control group. In 6-h pylorus-ligated animals, active fractions of drug at 100 mg/kg showed significant reduction in the ulcer index. Furthermore, in the pylorus-ligation model, significant reduction ( $p < 0.05$ ) was observed in total acidity, total acid output, pepsin activity, and pepsin output, along with a significant rise in the total carbohydrate to protein ratio (reflecting mucin activity) when compared with the control group. TLC studies revealed the presence of baicalein in the petroleum ether and hydrosylate in n.-butanol fraction. Fingerprinting of both the active fractions was developed by performing HPLC analysis. Baicalein was found to be a major flavonoid present both in petroleum ether and n.-butanol hydrosylate. The mechanism of its antiulcer activity could be attributed to a decrease in gastric acid secretory and antioxidant activities leading to gastric cytoprotection. This activity could be linked to the presence of baicalein in the root bark of the plant.

Keywords:

Antioxidant activity

antiulcer activity

gastric ulcer

*Oroxylum indicum*

## Introduction

*Oroxylum indicum*. Vent. (Bignoniaceae), commonly known as Syonakh, has been selected for the current study. This plant is used as an astringent, carminative, diuretic, stomachic, and aphrodisiac and is valued for stimulating digestion, curing fevers, coughs and other respiratory disorders (John, [2001](#)). *Oroxylum indicum*. is used as one of the important ingredients in the commonly used Ayurvedic preparation “dasamula.” It is also used in other Ayurvedic formulations such as amartarista, dantyadyarista, narayana taila, dhanawantara ghrita, brahma rasayana, chyavanaprasa awalwha, and so forth (Anonymous, [1998](#)). The plant is reported to possess anti-inflammatory, diuretic, anti-arthritis, antifungal, and antibacterial activities (Warrier et al., [1995](#)). The stem bark and leaves of this plant are reported to contain several flavonoids, namely, chrycin, oroxylin-A, scutellarin, and baicalein (Sankara et al., [1972a](#)[1972b](#)). Seeds of this plant are reported to contain ellagic acid (Vasantha et al., [1991](#)). However, the root

activity. Therefore, the current work aimed to study the effect of a 50% alcohol extract of *Oroxylum indicum*. and its different fractions on experimentally induced gastric ulcer models.

## Materials and Methods

### Procurement of plant material and extraction procedure

The fresh root bark of *Oroxylum indicum*. was collected in January 2005 from Vanaushadhi Ektrikaran Udyan, Ahwa, Dang Forest, Gujarat, India. The authentication of this plant was established by the taxonomist of Gujarat Ayurved University, Jamnagar, India, and a voucher specimen (404) was deposited in the Department of Pharmacognosy and Phytochemistry, L. M. College of Pharmacy, Ahmedabad, India. The root bark was sun-dried and powdered to 60 mesh. The powder of root bark after defatting with petroleum ether (0.32% w/w) was dried and then moistened with ammonia solution and extracted with chloroform (0.78% w/w), ethyl acetate (1.52% w/w), and n.-butanol (1.68% w/w), successively. The dried fractions were stored in an air-tight borosil glass container until further use.

### Drugs and chemicals

Omeprazole was obtained from Zydus Research Centre, Ahmedabad, India. All different organic solvents and reagents used for the current study were of analytical grade (AR) and obtained from S.D. Chem. Pvt. Ltd. (Mumbai, India). The standard baicalein was obtained from Sigma-Aldrich (St. Louis, MO, USA). Fresh drug solutions were prepared in 1% carboxy methylcellulose (CMC) and were administered orally.

### Animals

Wistar albino rats (Zydus Cadila Limited, Ahmedabad, India) of either sex weighing 175–225 g were selected for the current study. Animals were fed a standard chow diet and water that was freely available and maintained under standard conditions of a 12-h dark-light cycle,  $60\pm10\%$  humidity, and a temperature of  $21.5\pm1^{\circ}\text{C}$ . Coprophagy (and thus re-ingestion of any drug) was prevented by keeping the animals in cages with gratings on the floors. The distribution of animals in the groups, the sequence of trials, and treatment allotted to each group was randomized. Freshly prepared solutions of

university guidelines for animal experimentation. Throughout the entire study period, the rats were monitored for growth, health status, and food intake capacity to be certain that they were healthy. Utmost care was taken to ensure that animals were treated in the most humane and ethically acceptable manner. The animals were sacrificed with an overdose of ether anesthesia after the completion of the experiments. The stomachs were removed, opened along the greater curvature, washed with saline, and examined using a 6.4 binocular magnifier. Lesions were assessed by two unbiased observers.

## Methodology

The animals were divided into following groups of six.

- Group I (control): Rats received only aqueous suspension of 1% CMC vehicle with respect to the individual ulcerogenic procedure.
- Group II (drug treatment): Rats received the following treatments: 50% alcohol extract, petroleum ether, chloroform, ethyl acetate, and n.-butanol extracts (100-300 mg/kg, p.o.).
- Group III: Rats received standard omeprazole (20 mg/kg, p.o.) 1 h before the ulcerogenic procedure.

## Ethanol-induced gastric mucosal damage

Gastric lesions were induced by 1 ml absolute ethanol in 24-h fasted rats as per the method of Robert ([1979](#)). In the treatment group, drug extracts in 1% CMC solution were administered orally 1 h before the administration of ethanol. Animals were sacrificed 2 h after the ethanol administration, and gastric lesions were measured in terms of ulcer index (UI) determined by the method of Goswami et al. ([1997](#)). Each lesion of the stomach was measured along the greatest length and breadth. For circular lesions, the diameter was measured and area calculated. In case of petechies, five of them were considered to be equivalent to 1 mm<sup>2</sup> of ulcerated area. The total area of the stomach mucosa and that of ulcerated mucosa were calculated: Ulcer index = 10/X, where X = (Total mucosal area)/(Total ulcerated area). Further, the effect of drug administration on antioxidant enzymes and on lipid peroxide levels was evaluated. The stomach of each rat in each case was dissected out quickly, blood blotted off, washed with ice cold saline, and a 10% homogenate was prepared in phosphate buffer (10 mM,

C-24 high speed cooling centrifuge (Japan). The clear supernatant was used for the estimation of lipid peroxidation (MDA content), endogenous antioxidant enzymes (superoxide dismutase and catalase), and reduced glutathione. The assay for microsomal lipid peroxidation was carried out according to the method of Kiso et al. (1984). The superoxide dismutase (SOD) activity in the samples was determined the method of Mishra et al. (1973). Catalase (CAT) activity was measured according to the method of Aebi (1974). The reduced glutathione (GSH) was determined by the method of Beutler et al. (1963). The protein concentration in all samples was determined by the method of Lowry (1951).

## Pylorus-ligation (PL) model

Rats fasted for 24 h were anesthetized with ether, and a portion of abdomen was opened by a small midline incision below the xiphoid. The pylorus portion of the stomach was lifted and ligated (with care being taken not to occlude blood vessels) by the method of Shay et al. (1945). The stomach was closed with interupted sutures. Six hours after the pylorus ligation, animals were sacrificed. The stomach was dissected and the contents collected, measured, centrifuged, and subjected to biochemical analysis described below. Parameters investigated include: a ulcer index (UI) as described earlier, b acid secretory parameters, and c mucoprotective parameters. Acid secretory parameters include measurement of volume of gastric secretion, total acidity determined by titrating against 0.01 N sodium hydroxide to pH 8.0 using phenolphthalein as an indicator (Hawk et al., 1954), and total acid output (product of total acidity and volume of gastric secretion). Further, pepsin activity was determined using hemoglobin as a substrate, according to the modified method of Debnath et al. (1974). Total carbohydrates (TC) (Nair, 1976), total protein content (PR) (Lowry et al., 1951), mucin activity (TC/PR), and gastric mucus content (g) (Alarcon et al., 1993) were considered as a measure of the mucoprotective parameters.

## Fingerprinting and estimation of flavonoid

Both petroleum ether and n.-butanol fractions were found to be equipotent, showing a significant reduction in the extent of ulceration in both the models. This observation led us to the conclusion that a compound present in petroleum ether might also be present in n.-butanol possibly in glycosidal form. To confirm this, we hydrolyzed the n.-butanol fraction and performed a TLC study of both petroleum ether and hydrolysate of n.-

## TLC study of both active fractions

Hydrolysis of n.-butanol: 5 g of n.-butanol fraction was dissolved in water: methanol (9:1) solution hydrolyzed using 2 N HCl by refluxing the mixture for 2 h. After cooling, the ethyl acetate (0.64%) soluble fraction was separated and used for the further studies (TLC, HPLC). TLC co-chromatography was performed on the petroleum ether fraction, the hydrolyzed n.-butanol fraction, and standard baicalein.

### Method of TLC analysis

Ten microliters of each sample solution was spotted on the TLC plate (precoated with silica gel 60 F<sub>254</sub>, thickness 0.2 mm, 20 × 20 cm) (E Merck, Darmstadt, Germany) along with a standard solution of baicalein. Chromatogram was developed using chloroform:ethyl acetate:formic acid (10:8:2) as a mobile phase and visualized using natural product poly ethylene glycol (NP/PEG) reagent. On the basis of the TLC study, an HPLC method was developed for the quantification of baicalein in both the active fractions and development of fingerprints of the same.

### HPLC analysis

Chemicals: Methanol (HPLC grade), acetonitrile (HPLC grade), water (HPLC grade), trifluoroacetic acid (TFA) (analytical grade), baicalein (Sigma-Aldrich, Powai, Mumbai). HPLC was performed on a Shimadzu 2010 C (Tokyo, Japan), equipped with a C-18 Hypersil BDS column (250 × 4.6 mm, 5 µm). The instrument was operated under the following conditions: UV visible detector 254 nm, flow rate of 1.0 ml/min, retention time 40.5 min, injection volume 10 µl, and mobile phase A, water (pH = 2.70 adjusted with dilute H<sub>2</sub>SO<sub>4</sub>), mobile phase B, acetonitrile [diluent methanol: water pH = 3.0 with TFA (8:2)]. HPLC analysis of both the fractions was carried out for developing fingerprinting and also to verify the presence of baicalein in these extracts.

### Sample preparation for HPLC analysis

A calibration curve was obtained using standard solution of baicalein (1 mg/ml) at different concentrations in the range 25–150 µg/ml. Test solutions (1 mg/ml) of each extracts were prepared in methanol.

### Validation of HPLC method

The HPLC method was validated by determining precision, accuracy (% recovery), linearity, the linearity of calibration range, the limit of quantification, and the limit of detection.

## Statistical analysis

The results were expressed in terms of mean $\pm$ SEM. The significance of difference between mean values for the various treatments was tested using one-way analysis of variance test (ANOVA test) followed by Tukey's multiple range tests (Bolton, [1997](#)) wherever applicable to assess statistical significance of difference between the groups.

## Results

### Ethanol-induced gastric mucosal damage

Alcohol extract and the different fractions (300 mg/kg) showed a significant reduction in the ulcer index when compared with the control group, and results were comparable with the omeprazole-treated rats ([Table 1](#)). Reduction in the ulcer index was found to be maximum with both the n.-butanol (99.5%) and petroleum ether (96.0%) fractions at 100 mg/kg dose level as compared with control and omeprazole (99.5%) treatment ([Table 2](#)).

Table 1. Effect of different extracts (300 mg/kg, p.o.) of *Oroxylum indicum*. on ethanol-induced gastric mucosal damage in rats.

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Table 2. Effect of active fractions (100 mg/kg, p.o.) of *Oroxylum indicum*. on ethanol-induced gastric mucosal damage in rats.

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### Effect on free-radical activity

Petroleum ether and n.-butanol fractions showed significant reduction in MDA content when compared with the control group. In addition, significant increases in SOD, CAT, and reduced GSH levels were also observed, and the results were comparable with those of omeprazole treatment ([Table 3](#)).

Table 3. Effect of different extracts of *Oroxylum indicum*. (p.o.) on lipid peroxidation and antioxidant enzymes against ethanol-induced gastric mucosal damage.

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## Pylorus-ligation gastric ulcer model

The petroleum ether, n.-butanol fractions, and omeprazole pretreated rats showed significant reduction in the ulcer index when compared with the control group ([Table 4](#)).

Table 4. Effect of active fractions (100 mg/kg, p.o.) of *Oroxylum indicum*. on pylorus-ligated gastric ulcer model.

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## Effect on acid secretory parameters

Both the active fractions of drug and omeprazole treatment showed significant decrease in the volume of gastric secretion along with significant increase in the gastric pH, as compared with control group. They also showed significant reduction in total acidity, total acid output, pepsin activity, and pepsin output as compared with control group ([Table 5](#)).

Table 5. Effect of active fractions of *Oroxylum indicum*. (100 mg/kg, p.o.) on acid secretory parameters in pylorus-ligated gastric ulcer model.

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The petroleum ether and n.-butanol fractions showed significant reduction in the protein content of the gastric juice with no change in the carbohydrate content as compared with control group. Whereas, omeprazole treatment caused significant reduction in the TC and PR content, thus suggesting no improvement in the mucin activity. Therefore, TC:PR ratio (mucin activity) was significantly increased by both fractions. Furthermore, the gastric mucus content was found increased in petroleum ether and n.-butanol fractions pretreated animals as compared with control group. Omeprazole treatment also showed significant rise in mucus content of gastric mucosa (Table 6).

Table 6. Effect of active fractions of *Oroxylum indicum*. (100 mg/kg, p.o.) on mucoprotective parameters in pylorus-ligated gastric ulcer model.



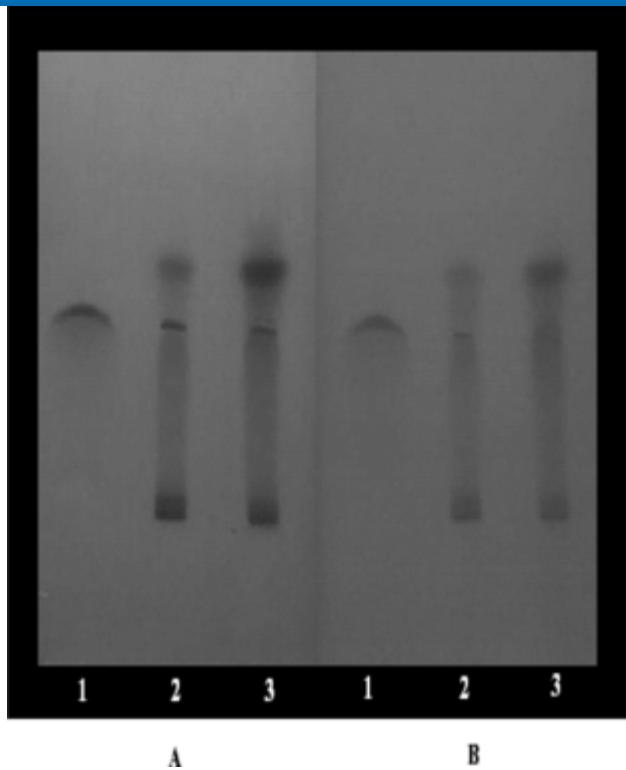
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## Fingerprinting and estimation of baicalein

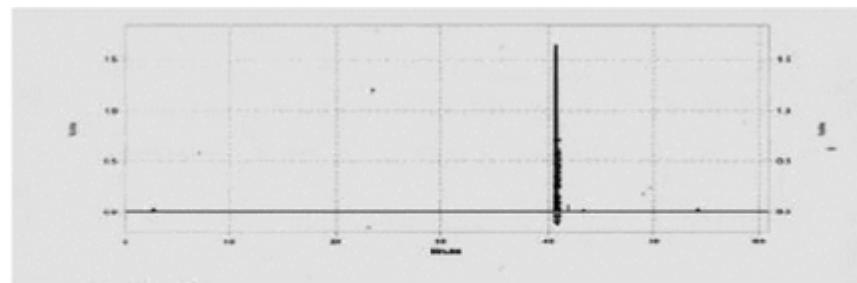
Based on the results of antiulcer activity, TLC study was aimed at checking the presence of baicalein in active fractions. Baicalein is reported to be present in stem bark and leaves of *Oroxylum indicum*. Our observations on TLC support the presence of baicalein, a major flavonoid (Fig. 1). Further, the authentic sample of baicalein resolved at 0.42 retention time ( $R_t$ ), and nearly the same  $R_t$  was observed with the use of petroleum ether and hydrolyzed n.-butanol fractions (Fig. 2; Table 7). The quantification and validation of baicalein was done by using a calibration curve prepared under same HPLC conditions (Fig 3; Table 8). Hydrolyzed n.-butanol fraction showed 12% of baicalein as compared with that of 5% of baicalein in petroleum ether fraction. This could be due to the glycosidic nature of baicalein imparting better absorption ability to the gut mucosa.

**Figure 1.** Co-chromatography of active fractions and standard baicalein. (A) Detected in UV (254 nm); (B) detected in UV (366 nm). 1, standard baicalein; 2, petroleum ether fraction; 3, hydrolyzed n.-butanol fraction.

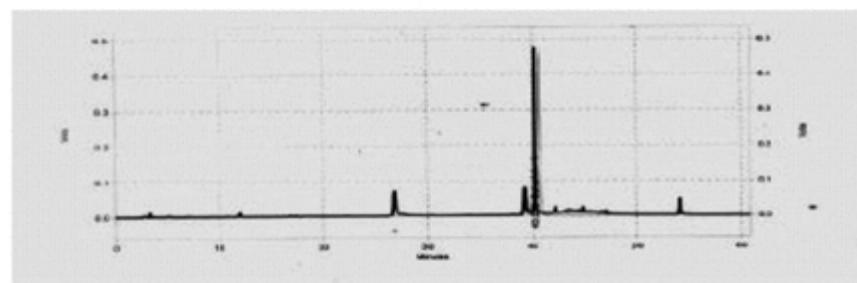


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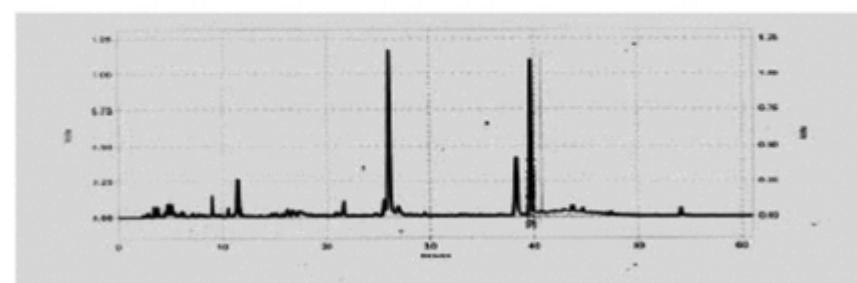
**Figure 2.** HPLC chromatograms of active fractions and standard baicalein. (a) Standard baicalein, (b) petroleum ether fraction, (c) hydrolyzed n.-butanol fraction.



a



b



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Figure 3. Calibration curve of standard baicalein.

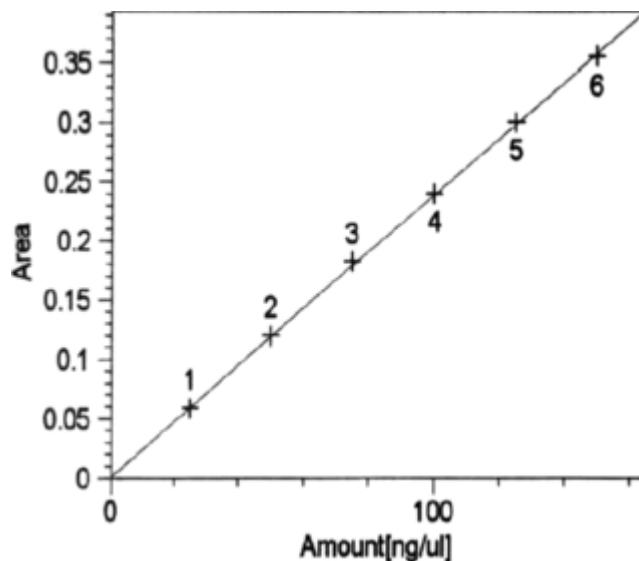
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Table 7. Percent of reference standard baicalein in active fractions of *Oroxylum indicum*.

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Table 8. Validation parameters of baicalein by HPLC.

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## Discussion

In the current study, it is suggested that petroleum ether and n.-butanol fractions of *Oroxylum indicum*. possess significant antiulcer activity as compared with omeprazole. Gastric mucosal damage induced by ethanol is reported to be due to mucosal

[1988](#)), and eventual injury (Bou-Abboud et al., [1988](#)). Reactive oxygen species are known to be involved in the pathogenesis of ethanol-induced gastric mucosal injury in vivo. (Pihan et al., [1987](#)). This causes damage to the cell and cell membranes (Fridovich, [1978](#)). Petroleum ether and n.-butanol fractions showed significant antiulcer activity in this model along with alteration in antioxidant enzyme status. Preventive antioxidants, superoxide dismutase (SOD), and catalase (CAT) are the first line of defense against reactive oxygen species (Halliwell, [1995](#)). In addition, reduced glutathione (GSH) is a major low-molecular-weight scavenger of free radicals in the cytoplasm and an important inhibitor of free radical-mediated lipid peroxidation (Piper & Stiel, [1986](#)). It was observed in our study that the drug pretreatment resulted in significant reduction in MDA content, along with significant rise in SOD, CAT, and reduced GSH levels, suggesting their efficacy in preventing free radical-induced damage. The mechanism of antiulcer activity in this model, therefore, can be attributed to the free radical scavenging activity of this drug that in turn might lead to gastric cytoprotection.

Gastric acid and pepsin are important factors for the formation of ulcers in pylorus-ligated rats (Shay et al., [1945](#)). Increased synthesis of nucleic acids and metabolism of carbohydrates and other compensatory mechanism could also be responsible for the ulceration due to pylorus ligation (Robert et al., [1984](#)). We observed significant reduction in total acidity and pepsin activity along with significant increase in the gastric pH in drug-treated animals. Besides, there was a significant rise in mucin activity and mucus content. Therefore, it is suggested that the fractions suppressed the gastric damage caused by aggressive factors and cause increase in defensive factors in terms of gastroprotection.

Preliminary screening using TLC revealed the presence of baicalein as one of the flavonoids in both petroleum ether and hydrolyzed n.-butanol fractions. RP-HPLC analysis was performed to develop complete chemoprofile and to quantify the baicalein in both the active fractions. Baicalein is reported to possess antioxidant activity (Ng et al., [2000](#), Alarcon et al., [1995](#)) and antiulcer activity (Kennouf et al., [2003](#)). Thus, the antiulcer action of n.-butanol and petroleum ether fractions may be attributed to baicalein, a flavonoid present in root bark of *Oroxylum indicum*.. Furthermore, it is evident from our observations that the n.-butanol fraction showed better protection than the petroleum ether fraction with reference to antisecretory and defense mechanisms.

## Conclusions

It is concluded that both the n.-butanol and petroleum ether fractions of Oroxylum indicum. possess significant antiulcer activity. There was an inhibitory effect on acid secretory mechanisms and free radical scavenging activity and a significant rise in gastric mucin activity. Further, with the help of HPLC-based profiling techniques, the antiulcer activity could be linked to a significant extent to the presence of baicalein in both fractions.

## Acknowledgment

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### Related Research Data

[Mechanism of Antihepatotoxic Activity of Glycyrrhizin, I: Effect on Free Radical Generation and Lipid Peroxidation](#)

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# References

1. Aebi H (1974): Catalase. In: Bergrenyer HV, ed., Methods in Enzymatic Analysis, 2nd ed. New York, Academic Press, 2: p. 673-678.

[Google Scholar](#)

2. Alarcon De La Lastra C, Lopez A, Motilva V (1993): Gastro protection and prostaglandin E2 generation in rats by *Dittrichia viscosa*.. *Planta Med* 59: 497-501. [\[INFOTRIEVE\]](#), [\[CSA\]](#)

[PubMed](#) [Web of Science ®](#) [Google Scholar](#)

3. Alarcon De La Lastra C, Martin MJ, Motilva V, Jimenez M, La Casa C, Lopez A (1995): Gastro protection induced by silymarin, the hepatoprotective principle of *Silybum marianum*. in ischaemia reperfusion mucosal injury: Role of neutrophils. *Planta Med* 61: 116-119. [\[INFOTRIEVE\]](#), [\[CSA\]](#)

[PubMed](#) [Web of Science ®](#) [Google Scholar](#)

4. Anonymous (1998): The Ayurvedic Pharmacopoeia of India. New Delhi, Government of India, Ministry of Health and Family Welfare, Department of Indian System of Medicine and Homeopathy, 3: pp. 209-210. [\[CSA\]](#)

[Google Scholar](#)

5. Beutler E, Duron O, Kelly B (1963): Reduced glutathione estimation. *J Clin Med* 61: 882-889. [\[CSA\]](#)

[PubMed](#) [Web of Science ®](#) [Google Scholar](#)

6. Bolton S (1997): Analysis of variance. In: Swarbrick J, ed., Pharmaceutical Statistics: Practice and Clinical Application, Basel, Marcel Dekker, Drug and Pharmaceutical Sciences Series, pp. 265-325.

[Google Scholar](#)

7. Bou-Abboud CF, Wayland H, Paulsen G, Guth PH (1988): Micro-circulatory stasis

8. Debnath PK, Gode KD, Govinda D, Sanyal AK (1974): Effect of propanolol on gastric secretion in albino rats. *Br J Pharmacol* 51: 213-216. [INFOTRIEVE], [CSA]

9. Fridovich I (1978): The biology of free radicals. *Science* 201: 875-880. [INFOTRIEVE], [CSA]

10. Goswami S, Jain S, Santani D (1997): Anti-ulcer activity of cromakalin (BRL 34915), a potassium channel openers against experimentally induced gastric and duodenal ulcers in rats and guinea-pigs. *J Pharm Pharmacol* 49: 195-199. [INFOTRIEVE], [CSA]

11. Halliwell B (1995): Antioxidant characterization: Methodology and mechanism. *Biochem Pharmacol* 49: 1341-1345. [INFOTRIEVE], [CSA], [CROSSREF]

12. Hawk P, Oser B, Summerson W (1954): Gastric analysis. In: Stevens E, ed., *Practical Physiological Chemistry*. Toronto, Blakiston, pp. 378-380.

Google Scholar

13. John AP (2001): Healing Plants of Peninsular India.. In: Bignoniaceae, *Oroxylum indicum*. WallingfordUK, CABI publishing, p. 169-171.

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14. Kennouf S, Benabdallah H, Gharzouli K, Amira S, Ito H, Kim TH, Yoshida T, Gharzouli A (2003): Effect of tannins from *Quercus suber*. and species leaves on ethanol induced gastric lesions in mice. *J Agri Food Chem* 51: 1469-1473. [CSA], [CROSSREF]

5. Kiso Y, Tohkin H, Hikino H, Hattori M, Sakamoto T, Namba T (1984): Mechanism of antihepatotoxic activity of glycyrrhizin, I: Effect on free radical generation and lipid peroxidation. *Planta Med* 50: 298-302. [\[INFOTRIEVE\]](#), [\[CSA\]](#)

[PubMed](#) [Web of Science ®](#) [Google Scholar](#)

6. Lowry OH, Rosenbrough NJ, Farr AL, Randall RJ (1951): Protein measurement with Folin phenol reagent. *J Biol Chem* 193: 265-275. [\[INFOTRIEVE\]](#), [\[CSA\]](#)

[PubMed](#) [Web of Science ®](#) [Google Scholar](#)

7. Mishra HP, Frodovich I (1973): The role of super oxide anion in the auto oxidation of epinephrine and a simple assay for superoxide dismutase. *J Biol Chem* 247: 3170-3175. [\[CSA\]](#)

[Web of Science ®](#) [Google Scholar](#)

8. Nair BR (1976): Investigations on the venom of south Indian scorpion. *Heterometrus scaber*. Ph.D. thesis, University of Kerala, Trivendrum, p. 39.

[Google Scholar](#)

9. Ng TB, Lin F, Wang ZT (2000): Antioxidant activity of natural products from plants. *Life Sci* 68: 709-723. [\[CSA\]](#), [\[CROSSREF\]](#)

[PubMed](#) [Google Scholar](#)

20. Oates PJ, Hakkinen JP (1988): Studies on the mechanism ethanol-induced gastric damage in rats. *Gastroenterology* 94: 9-21. [\[CSA\]](#)

[Web of Science ®](#) [Google Scholar](#)

21. Peskar BM, Lange K, Hoppe U, Peskar BA (1986): Ethanol stimulates formation of leukotriene C4 in rat gastric mucosa. *Prostaglandins* 31: 283-293. [\[INFOTRIEVE\]](#), [\[CSA\]](#), [\[CROSSREF\]](#)

[PubMed](#) [Web of Science ®](#) [Google Scholar](#)

22. Bihari C, Boccellino C, Szabo S (1987): Free radicals and lipid peroxidation in ethanol- or

23. Piper DW, Stiel DD (1986): Pathogenesis of chronic peptic ulcer, current thinking and clinical implication. *Med Prog* 2: 7-9. [CSA]

[Google Scholar](#)

24. Robert A (1979): Cytoprotection by prostaglandins in rats-prevention of gastric necrosis produced by alcohol, HCl, NaOH, hypertonic NaCl and thermal injury. *Gastroenterology* 77: 433-443. [INFOTRIEVE], [CSA]

25. Robert A, Bottcher W, Golanska E, Kauffman GL (1984): Lack of correlation between mucus gel thickness and gastric cytoprotection in rats. *Gastroenterology* 86: 670-674. [INFOTRIEVE], [CSA]

26. Sankara S, Nair AGR (1972a): Flavonoids of the stem bark of *Oroxylum indicum*.. *Curr Sci* 41: 62-63. [CSA]

[Web of Science ®](#) [Google Scholar](#)

27. Sankara S, Nair AGR (1972b): Flavonoids from the leaves of *Oroxylum indicum*. and *Pajanelia longifolia*.. *Phytochemistry* 11: 439-440. [CSA], [CROSSREF]

28. Shay H, Komarov SA, Fels SS, Meraze D, Gruenstein M, Siplet H (1945): A simple method for the uniform production of gastric ulceration in rats. *Gastroenterology* 5: 43-61. [CSA]

[Web of Science ®](#) [Google Scholar](#)

29. Vasantha S, Natarajan M, Suderesan R, Bhima Rao R, Kundu AB (1991): Ellagic acid from root bark of *Oroxylum indicum*.. *Indian Drugs* 28: 507. [CSA]

30. Warrier PK, Nambiar VPK, Ramankutty C (1995): *Oroxylum indicum*.. In: A Compendium of 500 Species, Indian Medicinal Plants, Vol IV. Madras, Orient Longman Ltd., pp. 186–190.

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