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Author's Views

SESTRINs regulate mTORC1 via RRAGs: The riddle of GATOR

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Abstract

SESTRINs, proteins encoded by the SESN1–3 genes in mammals, are well-established suppress[illegible] kinase. Recently [illegible] is a regulato[illegible] support the RRAGA/B[illegible] of GATOR2 in this p[illegible]

factors, and stress to control metabolism, autophagy, and cell growth. mTORC1 activity inversely correlates with the lifespan of most eukaryotes and mTORC1 contributes to several different pathologies including cancer, diabetes, and neurodegenerative diseases.[2] mTORC1 is activated by 2 groups of small guanine triphosphatases (GTPases): Ras homolog enriched in brain (RHEB) and members of the Ras-related GTP-binding protein (RRAG) family, working as RRAG-A/B and RRAG-C/D heterodimers. RHEB activity is inhibited by the tuberoses sclerosis protein complex (TSC). TSC in turn is controlled by several stress insults via activation of 5´-AMP-activated protein kinase (AMPK), and we have shown previously that SESTRINs inhibit mTORC1 via the AMPK-TSC axis.[3] Amino acids and glucose control mTORC1 via RRAGs, which stimulate translocation of mTORC1 to the lysosomes where it can be activated by RHEB. RRAG-A/B is activated by its GTPase exchange factor (GEF) called ragulator and is inhibited by the protein complex GATOR1 (GTPase activating protein [GAP] activity toward RRAG complex 1) working as a GAP. GATOR1 in turn is suppressed by the GATOR2 protein complex within the GATOR supercomplex.[4]

Three groups recently reported a novel mechanism of mTORC1 inhibition by SESTRINs via suppression of RRAG-dependent mTORC1 lysosomal translocation.[5-7] SESTRINs are critical for mTORC1 inhibition by amino acid withdrawal in mammalian cells. Surprisingly, SESTRINs do not affect RRAG-A/B GDP/GTP loading, suggesting that they are not regulators of RRAG-A/B GAP or GEF. Although all 3 groups agreed on the importance of RRAGs for mTORC1 suppression by SESTRINs, they proposed different mechanisms for this regulatory activity.

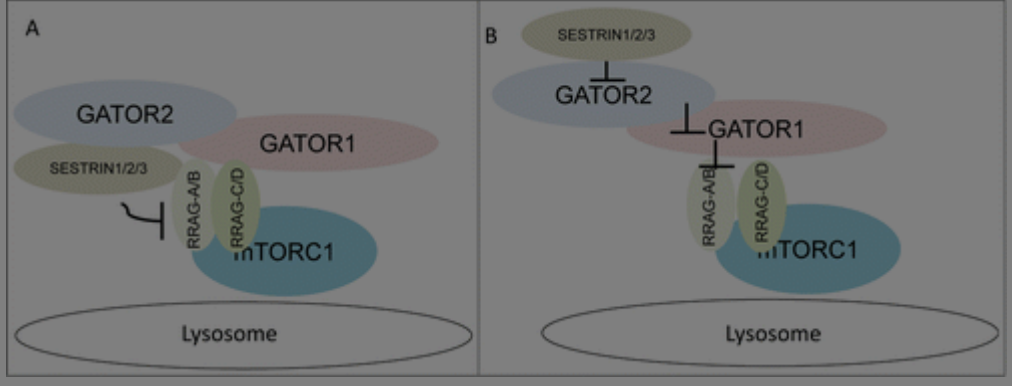
Using m omplexes, our grou time, Sabatini ng similarity to our da a GATOR- and RRA ATOR2 interacti way from GAT

Li's grou INs.[6] Accordin rk as RRAG-A/B guar shares homolog GTPase

family members. Mutations in the GDI motif significantly impair the ability of SESTRIN2 to inhibit mTORC1 and delivery of a peptide GDI motif is sufficient to inhibit mTORC1.[6]

There are several major concerns that remain unexplained. First, do SESTRINs directly interact with RRAGs and what is the role of the GATOR complex in the regulation of RRAG-A/B and mTORC1 activities by SESTRINs? Despite the high abundance of GATOR2 peptides among our mass spectrometry data, we did not observe any RRAG peptides, indicating that this interaction is very weak and might be indirect. Second, according to Li's model, SESTRIN2 must have some regulatory role over RRAGs even in the absence of GATOR1 as a result of intrinsic RRAG-A/B GTPase activity,[6] although data from our group and Sabatini's group indicate a strong requirement for the GATOR complex in the regulation of mTORC1 by SESTRINs.[5,7] Finally, in contrast to Peng et al., we were not able to detect any colocalization of SESTRIN2 with lysosomes, indicating that SESTRINs might play a major role in the regulation of GATOR and RRAGs in the cytoplasm.[6,7] To reconcile these studies, we propose that GATOR is required for the interaction between SESTRIN2 and RRAGs, potentially providing a bridge between them and somehow bringing the potential GDI domain of SESTRINs into close proximity of the RRAG-A/B protein. Thus, GATOR might control interactions between SESTRINs and RRAGs and assist in mTORC1 inhibition by SESTRINs (Fig. 1A). Alternatively, SESTRINs might interact solely with GATOR2, and the GDI activity of SESTRIN toward RRAGA/B might be mediated by GATOR (Fig. 1B). We demonstrated that the interaction between SESTRIN2 and GATOR2 is mediated by WD-repeat domain 24 (WDR24) and SEH1-like (SEH1L) proteins, thus it would be important to examine whether the GDI domain of SESTRIN is required for the interaction between SESTRINs and GATOR2.

Figure 1. GATOR and RRAG-A/ e mTORC1 activity ulation of RRAG-A/ weakly with RRAG-A/ actions betw TRINs on RRAG-A/ ly mediated by GATO -A/B.



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To reconcile the discrepancy in localization studies we propose that the interaction between SESTRINs and GATOR–RRAGs complexes is a dynamic process and under certain conditions SESTRINs can be associated with lysosomes, although most of the proteins associated with GATOR are still located in the cytoplasm in close proximity to lysosomes. Our findings are consistent with recently published data from Avruch's laboratory[8] showing a predominantly cytoplasmic localization of RRAGs based on protein fractionation studies. In the future it would be important to analyze the localization of endogenous SESTRINs and RRAGs, and the role of SESTRINs in the control of RRAG localization in the cell under normal and stress conditions.

Interestingly, yeast mTORC1 is regulated by amino acids via a mechanism mediated by Gtr1 and Gtr2, orthologs of mammalian RRAG-A/B and RRAG-C/D, and the SEA complex (SEAC), an analog of mammalian GATOR. SEAC functions as a whole complex, suggesting that GATOR might also work as an entire complex to regulate RRAG-A/B activity.[9,10] However, the fact that yeasts do not express sestrins indicates that they are not oblig legans and D. melan lerated aging ph s rather than function y of SESTRIN amily might play dist nt SESTRINs migh rolled mTORC1 mage and switchin

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No potential conflicts of interest were disclosed.

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