

Introduction

Dexibuprofen (Dex), a novel non-steroidal anti-inflammatory drug (NSAID), is a single and pharmacologically effective enantiomer of racemic ibuprofen. Racibuprofen and dexibuprofen differ in their physicochemical properties, pharmacological profiles and metabolic activities¹. The efficacy of dexibuprofen was found to be same as that of other NSAIDs, such as diclofenac, naproxen and celecoxib. Generally, the NSAIDs are associated with various gastrointestinal (GI) side effects like stomach ulceration, bleeding and perforation due to the presence of its acidic group. The GI toxicity is produced either by direct contact mechanism or by generalized systemic action, which occur after absorption. To overcome the GI side effects, prodrugs of NSAIDs can be developed by hiding its acidic group through conjugation with amino acids. Many amino acids possess site specificity, marked anti-inflammatory activity and exhibit profound healing effect on gastric toxicity². During conjugation, the NSAIDs having free carboxylic group can be derivitized into corresponding amides of amino acid, and it results in altering the physical properties of parent drug with one or more of hydrolase enzymes serving as the in vivo reconversion sites. The new prodrugs were expected to have better lipophilicity, reduced gastric irritancy, improved therapeutic index through the prevention of GI irritation and bleeding, improved anti-inflammatory activity, reduced GI toxicity, etc.

The synthesis of prodrugs of dexibuprofen, etc., with amino acids like glycine, methyl ester, etc., has been reported so far. The objective of this study was to synthesize methyl ester, Dex-1, Dex-2, etc., with various amino acids. The physicochemical properties of these compounds were studied. The pharmacokinetic and pharmacodynamic studies were carried out in rats. The results are discussed in this article.

Materials and methods

Materials and methods

Materials

Results and discussion

The amino acids L-tryptophan, L-phenylalanine, glycine and L-tyrosine were obtained

from Mys Hi-Media Ltd., India, and drug dexibuprofen was obtained as a gift sample

from Alkem Laboratories, India. Other reagents and solvents were of analytical grade.

The melting points were recorded using melting point determination apparatus by

References

Sigma Instrument, India, and are uncorrected. The elemental analysis was performed

using Carlo-Erba Model 1108 Analyzer (Italy). ^1H NMR and ^{13}C NMR spectra were

recorded in dimethylsulphoxide (DMSO) on NMR spectrophotometer (Bruker DRX 300,

USA). Chemical shifts are expressed as δ (ppm) values. IR spectra were recorded using

IR spectrophotometer (Shimadzu FTIR-8201PC (Kyoto, Japan)) in KBr phase and mass

spectra were recorded on mass spectrophotometer (Jeol SX-102 (FAB), Japan). The

hydrolysis data and drug content determination were performed by ELICO Double Beam

UV-VIS Spectrophotometers (Hyderabad, India).

Synthesis of amide prodrugs of dexibuprofen

Dexibuprofen is 2-[4-(2-methyl propyl) phenyl] propanoic acid and the synthesis of its amide prodrugs was carried out by Schotten Baumann technique⁸ as explained below.

The purity of the compounds was checked by thin-layer chromatography on pre-coated silica GF₂₅₂ plates using iodine vapour as a detecting agent. The structures of all the synthesized prodrugs were confirmed by elemental analysis and spectral analysis such as IR, ^1H NMR, ^{13}C NMR and mass spectroscopy.

Step 1:

Dex (1) g was dissolved in 10 ml of dichloromethane (DCM) and freshly distilled triethylamine (TEA) was added. The mixture was refluxed for 30 min. The mixture became viscous yellow liquid. The colour of the mixture was yellow.

Step 2:

phenylpropanoic acid, L-

Freshly distilled ethanol (100 ml) was added to the mixture.

mixture was refluxed for 6–8 h at 60–70°C with continuous stirring on magnetic stirrer. Excess thionyl chloride and solvent was removed under reduced pressure giving crude tryptophan methyl ester hydrochloride. It was treated with 20 mL portion of cold ether at 0°C. The resulting solid product was collected and dried under vacuum. It was recrystallized from hot methanol by slow addition of 15–20 mL ether followed by cooling at 0°C. The crystals were collected next day and washed twice with ether: methanol mixture at 5:1 ratio followed by pure ether and dried under vacuum to give pure tryptophan methyl ester hydrochloride (2a). The same procedure was followed to synthesize phenylalanine methyl ester hydrochloride, glycine methyl ester hydrochloride and tyrosine methyl ester hydrochloride.

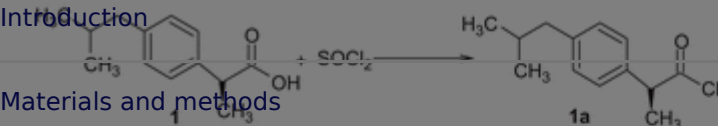
Step 3: Synthesis of prodrugs of dexibuprofen with methyl esters of L-tryptophan, L-phenylalanine, glycine and L-tyrosine

Ice cold, aqueous sodium hydroxide solution (5 %) was taken in 250 mL beaker and 12.6 g (0.05 mol L^{-1}) of methyl ester of tryptophan hydrochloride was added to it. The reaction mixture was mechanically stirred for 30 min at room temperature, after which the beaker was transferred to an ice bath kept on the mechanical stirrer, maintaining the temperature at 10°C. To this, 22.3 g (0.01 mol L^{-1}) of dex acid chloride was added in small portions with continuous stirring for 7–8 h. The solid that separated out was filtered off. The crude prodrug was recrystallized from methanol. The same procedure was followed for L-phenylalanine, glycine and L-tyrosine. The schematic representation of synthesis of tryptophan-conjugated dexibuprofen (Dex 1), phenylalanine-conjugated dexibuprofen (Dex 2), glycine-conjugated dexibuprofen (Dex 3) and tyrosine-conjugated dexibuprofen (Dex 4) is shown in Scheme 1.

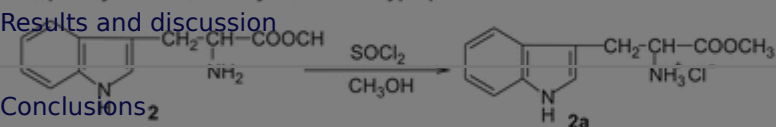
Scheme 1



Step I: Synthesis of dexibuprofen acid chloride

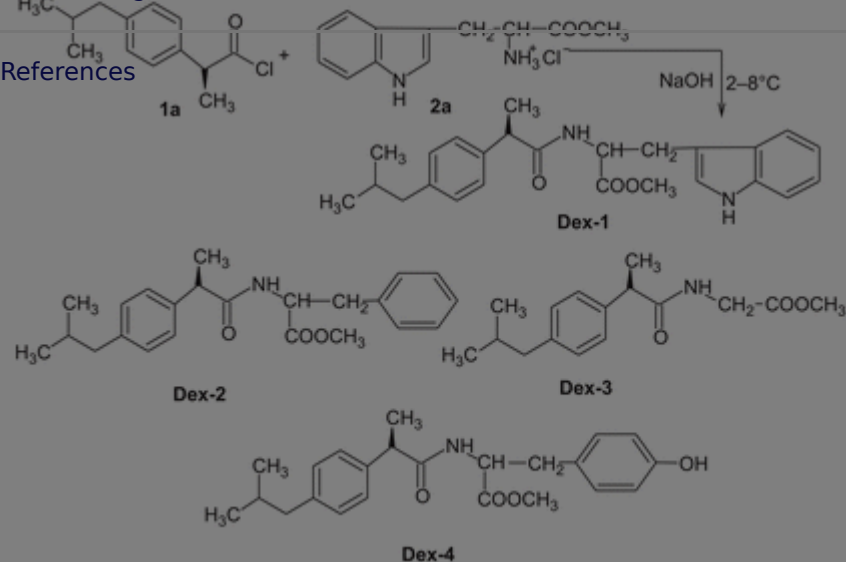


Step II: Synthesis of methyl ester of L-tryptophan



Step III: Synthesis of prodrug: Conjugation of dexibuprofen acid chloride with methyl ester of L-tryptophan

Acknowledgements



Display full size

Spectral analysis

The spectral data obtained for Dex 1, Dex 2, Dex 3 and Dex 4 are given as follows.

Dex 1: S(+)- Methyl 3-(1H-indol-3-yl)-2-(2-(4-isobutylphenyl)propanamido)propanoate—

UV Spectra (λ_{max}) in simulated gastric fluid (SGF) 205 nm, in simulated intestinal fluid (SIF) 225 nm, in 90% human plasma 219 nm; IR (KBr, cm^{-1}) 3300 (NH str. of amide),

3075 (aromatic C-H), 2950 (aliphatic C-H), 1735 (C=O str. of ester); 1H

NMR (δ) 7.25 (m, 1H, H-7), 7.15 (m, 1H, H-5), 7.05 (m, 1H, H-4), 6.95 (m, 1H, H-6), 3.72

(d, $J = 7.1$ Hz, 2H, CH₂), 9.77 (s, 1H, NH), 3.85 (s, 3H, OCH₃), 2.85 (s, 3H, CH₃), 2.35 (s, 3H, CH₃), 1.85 (s, 3H, CH₃), 1.35 (s, 3H, CH₃).

^{13}C NMR (δ) 173.5, 172.5, 171.5, 170.5, 169.5, 168.5, 167.5, 166.5, 165.5, 164.5, 163.5, 162.5, 161.5, 160.5, 159.5, 158.5, 157.5, 156.5, 155.5, 154.5, 153.5, 152.5, 151.5, 150.5, 149.5, 148.5, 147.5, 146.5, 145.5, 144.5, 143.5, 142.5, 141.5, 140.5, 139.5, 138.5, 137.5, 136.5, 135.5, 134.5, 133.5, 132.5, 131.5, 130.5, 129.5, 128.5, 127.5, 126.5, 125.5, 124.5, 123.5, 122.5, 121.5, 120.5, 119.5, 118.5, 117.5, 116.5, 115.5, 114.5, 113.5, 112.5, 111.5, 110.5, 109.5, 108.5, 107.5, 106.5, 105.5, 104.5, 103.5, 102.5, 101.5, 100.5, 99.5, 98.5, 97.5, 96.5, 95.5, 94.5, 93.5, 92.5, 91.5, 90.5, 89.5, 88.5, 87.5, 86.5, 85.5, 84.5, 83.5, 82.5, 81.5, 80.5, 79.5, 78.5, 77.5, 76.5, 75.5, 74.5, 73.5, 72.5, 71.5, 70.5, 69.5, 68.5, 67.5, 66.5, 65.5, 64.5, 63.5, 62.5, 61.5, 60.5, 59.5, 58.5, 57.5, 56.5, 55.5, 54.5, 53.5, 52.5, 51.5, 50.5, 49.5, 48.5, 47.5, 46.5, 45.5, 44.5, 43.5, 42.5, 41.5, 40.5, 39.5, 38.5, 37.5, 36.5, 35.5, 34.5, 33.5, 32.5, 31.5, 30.5, 29.5, 28.5, 27.5, 26.5, 25.5, 24.5, 23.5, 22.5, 21.5, 20.5, 19.5, 18.5, 17.5, 16.5, 15.5, 14.5, 13.5, 12.5, 11.5, 10.5, 9.5, 8.5, 7.5, 6.5, 5.5, 4.5, 3.5, 2.5, 1.5, 0.5.

43.5, 47.5, 48.5, 49.5, 50.5, 51.5, 52.5, 53.5, 54.5, 55.5, 56.5, 57.5, 58.5, 59.5, 60.5, 61.5, 62.5, 63.5, 64.5, 65.5, 66.5, 67.5, 68.5, 69.5, 70.5, 71.5, 72.5, 73.5, 74.5, 75.5, 76.5, 77.5, 78.5, 79.5, 80.5, 81.5, 82.5, 83.5, 84.5, 85.5, 86.5, 87.5, 88.5, 89.5, 90.5, 91.5, 92.5, 93.5, 94.5, 95.5, 96.5, 97.5, 98.5, 99.5.

129.5, 128.5, 127.5, 126.5, 125.5, 124.5, 123.5, 122.5, 121.5, 120.5, 119.5, 118.5, 117.5, 116.5, 115.5, 114.5, 113.5, 112.5, 111.5, 110.5, 109.5, 108.5, 107.5, 106.5, 105.5, 104.5, 103.5, 102.5, 101.5, 100.5, 99.5, 98.5, 97.5, 96.5, 95.5, 94.5, 93.5, 92.5, 91.5, 90.5, 89.5, 88.5, 87.5, 86.5, 85.5, 84.5, 83.5, 82.5, 81.5, 80.5, 79.5, 78.5, 77.5, 76.5, 75.5, 74.5, 73.5, 72.5, 71.5, 70.5, 69.5, 68.5, 67.5, 66.5, 65.5, 64.5, 63.5, 62.5, 61.5, 60.5, 59.5, 58.5, 57.5, 56.5, 55.5, 54.5, 53.5, 52.5, 51.5, 50.5, 49.5, 48.5, 47.5, 46.5, 45.5, 44.5, 43.5, 42.5, 41.5, 40.5, 39.5, 38.5, 37.5, 36.5, 35.5, 34.5, 33.5, 32.5, 31.5, 30.5, 29.5, 28.5, 27.5, 26.5, 25.5, 24.5, 23.5, 22.5, 21.5, 20.5, 19.5, 18.5, 17.5, 16.5, 15.5, 14.5, 13.5, 12.5, 11.5, 10.5, 9.5, 8.5, 7.5, 6.5, 5.5, 4.5, 3.5, 2.5, 1.5, 0.5.

Dex 2: S(+)- Methyl 3-(1H-indol-3-yl)-2-(2-(4-isobutylphenyl)propanamido)propanoate—UV

Spectra (λ_{max}) in simulated gastric fluid (SGF) 205 nm, in simulated intestinal fluid (SIF) 225 nm, in 90% human plasma 219 nm; IR (KBr, cm^{-1}) 3300 (NH str. of amide),

3075 (aromatic C-H), 2950 (aliphatic C-H), 1735 (C=O str. of ester), 1715 (C=O str. of amide), 1600 (C=C str. of aromatic ring), 1500 (C-N str. of amide), 1450 (C-O str. of ester), 1350 (C-O str. of ester), 1250 (C-O str. of ester), 1150 (C-O str. of ester), 1050 (C-O str. of ester), 950 (C-O str. of ester), 850 (C-O str. of ester), 750 (C-O str. of ester), 650 (C-O str. of ester), 550 (C-O str. of ester), 450 (C-O str. of ester), 350 (C-O str. of ester), 250 (C-O str. of ester), 150 (C-O str. of ester).

1H NMR (δ) 7.25 (m, 1H, H-7), 7.15 (m, 1H, H-5), 7.05 (m, 1H, H-4), 6.95 (m, 1H, H-6), 3.72 (d, $J = 7.1$ Hz, 2H, CH₂), 9.77 (s, 1H, NH), 3.85 (s, 3H, OCH₃), 2.85 (s, 3H, CH₃), 2.35 (s, 3H, CH₃), 1.85 (s, 3H, CH₃), 1.35 (s, 3H, CH₃).

4H, Ar, H), 7.32 (m, 4H, Ar. ring), 9.77 (s, 1H, NH); ¹³C NMR (δ ppm) (DMSO-d₆) 17.3, 17.3, 18.3, 35.2, 35.2, 42.3, 47.3, 47.8, 51.9, 119.2, 120.2, 125.2, 125.3, 125.3, 126.2, 126.6, 127.2, 127.3, 127.5, 128.3, 128.4, 171.3, 172.9; Mass (m/z) 367 (M+).

Results and discussion

Dex 3: S(+)-Methyl-2-(2-(4-isobutyl phenyl)propanamido)acetate—UV Spectra (λ_{max}) in SGF 212 nm, in SIF 219 nm, in 80% human plasma 222 nm; IR (KBr, cm⁻¹) 3280 (NH str.

of amide), 3043 (aromatic CH str.), 2932 (C=O str. of ester), 1630 (C=N), 1290 (CO str. of ester); ¹H NMR (δ ppm) (DMSO-d₆) 1.38 (d, J = 6.7 Hz, 6H, CH₃), 2.42 (d, J = 7.3 Hz, 2H, CH₂), 3.80 (q, J = 7.1 Hz, 1H, CH), 3.82 (q, J = 7.1 Hz, 1H, CH), 7.48 (d, J = 8 Hz, 4H, Ar.), 9.68 (s, 1H, NH); ¹³C NMR (δ ppm) (DMSO-d₆) 17.3, 17.3, 17.8, 20.2, 35.2, 42.3, 47.8, 51.9, 63.2, 120, 120.8, 128.6, 128.9, 139.6, 172.9; Mass (m/z) 277 (M+).

Dex 4: S(+)-Methyl 3-(4-hydroxyphenyl)-2-(2-(4-isobutylphenyl)propanamido)propanoate—UV Spectra (λ_{max}) in SGF 208 nm, in SIF 221 nm, in 80 % human plasma 218 nm; IR (KBr, cm⁻¹) 3362 (NH str. of amide), 3046 (aromatic CH str.), 1745 (C=O str. of ester), 1532 (C=N), 1280 (CO str. of ester); ¹H NMR (δ ppm) (DMSO-d₆) 1.28 (d, J = 6.7 Hz, 6H, CH₃), 3.62 (q, J = 7.1 Hz, 1H, CH), 7.28 (d, J = 8 Hz, 4H, Ar. ring), 7.63 (d, J = 8 Hz, 4H, Ar. ring), 9.68 (s, 1H, NH); ¹³C NMR (δ ppm) (DMSO-d₆) 17.8, 18.2, 18.3, 36.2, 47.3, 48.2, 48.3, 51.9, 120, 120.2, 120.6, 122, 122.2, 125, 125.6, 126, 126.2, 128.2, 128.4, 135.2, 171.2, 172.1; Mass (m/z) 383 (M+).

Physicochemical characterization of the synthesized prodrugs

Solubility

Approximately 10 mg of each prodrug was dissolved in 10 mL of distilled water at ± 1°C in glass test tubes. The solutions were then diluted with 10 mL of ethanol, ether, and chloroform. The solutions were gently shaken and the resulting suspensions were known as the prodrug suspensions. The compounds were then used for the study.



Protein

A solution of 10 mg of prodrug in 10 mL of distilled water was offered saline (PBS, pH 7.4) to mice. The prodrug was then administered from Hi-

7.4). It was tied at the opening end of dialysis tube; the dialysis tube containing (6%) egg albumin was dipped into the drug solution and covered. The whole assembly was placed on a magnetic stirrer and switched at low revolutions per minute. The temperature was maintained at $37 \pm 0.5^\circ\text{C}$. After every 1 h, 1 mL of the PBS containing drug solution was replaced with fresh 1 mL of PBS. Withdrawn sample was diluted further with 1 mL phosphate buffer and the concentration of prodrug was estimated using spectrophotometer at 223 nm. Table 1 indicates the physicochemical properties of the synthesized prodrugs.

Table 1. Physicochemical properties of prodrugs.

Download CSV

Display Table



Hydrolysis rate determination in SGF and SIF⁹

In vitro hydrolysis studies of synthesized prodrugs were carried out in SGF at pH 1.2 (USP 1970) and SIF at pH 7.4 (USP 1970). A solution of 10 mg of prodrug was prepared in 90 mL of SIF (pH 7.4) or SGF (pH 1.2). An aliquot of 15 mL of this solution was withdrawn repeatedly and kept in test tubes maintained at $37 \pm 0.5^\circ\text{C}$. At a definite interval of time (0.5 h, 1-8 h), an aliquot was withdrawn from different test tubes and was transferred to micro-centrifuge tubes followed by addition of methanol to make up the volume. The tubes were placed in freezing mixture in order to arrest further hydrolysis, followed by vortexing at high speed for 5 min. After vortexing, the tubes

were centrifuged and the amount obtained from each test tube was determined. The amount of free Dex released from each test tube was determined. The kinetics of hydrolysis was studied by plotting the amount of prodrug and order of reaction. The rate constant was calculated using the following equation:

$$r = (2.303) \log \frac{C_0}{C_t}$$

where r is the rate of hydrolysis, C_0 is the initial concentration of prodrug, C_t is the concentration of prodrug remaining at time t .

Introduction
A solution of 10 mg of prodrug was prepared in methanol (2 mL) and was added to 88 mL of 80% human plasma (pH 7.4 prepared by mixing 80 portion of plasma and 20 portion of phosphate buffer at pH 7.4). An aliquot of 15 mL of this solution was withdrawn and kept in test tubes maintained at $37 \pm 0.5^\circ\text{C}$. At definite interval of time (0-8 h), an aliquot of 22 mL was withdrawn and mixed with 0.5 % ZnSO_4 solution.

Conclusions
Samples were centrifuged at 6000g for 10 min and a clear supernatant solution was analyzed spectrophotometrically at 223 nm.

References

Hydrolysis rate determination in rat faecal matter¹¹

The prodrug was dissolved in phosphate buffer so that final concentration of the solution was 250 $\mu\text{g/mL}$. Fresh faecal material of rats were weighed (about 1 g) and placed in different sets of test tubes. To each test tube, 2 mL of the prodrug solution was added and diluted to 5 mL with phosphate buffer. The sets of test tubes were incubated at $37 \pm 0.5^\circ\text{C}$ for different intervals of time (0-8 h). For analysis, the free drug was extracted with 5 mL of methanol and estimated on UV spectrophotometer at 210 nm.

Pharmacological evaluations

Dex and the synthesized prodrugs were evaluated for anti-inflammatory activity, analgesic activity, ulcerogenicity and histopathology, and a comparative study was performed. Test compounds and standard drugs were administered in the form of a suspension by oral route of administration for anti-inflammatory and analgesic studies (1% carboxymethylcellulose as a vehicle), as well as for ulcerogenicity studies (2%

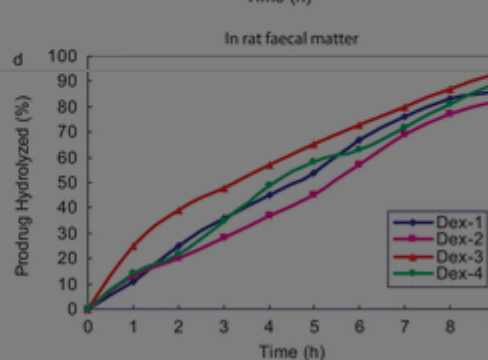
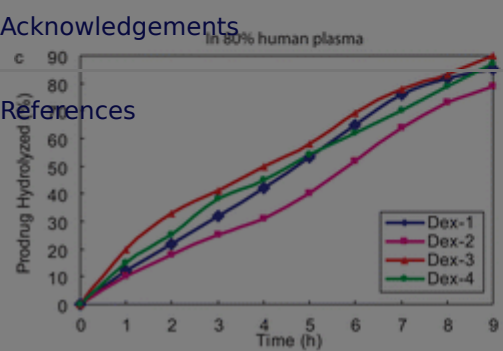
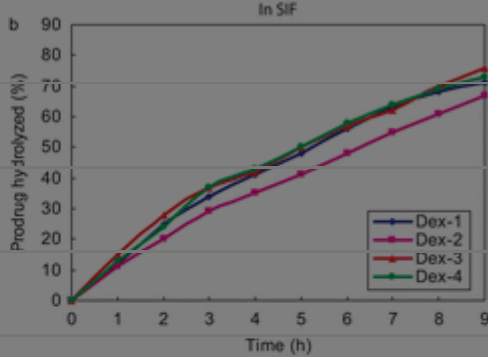
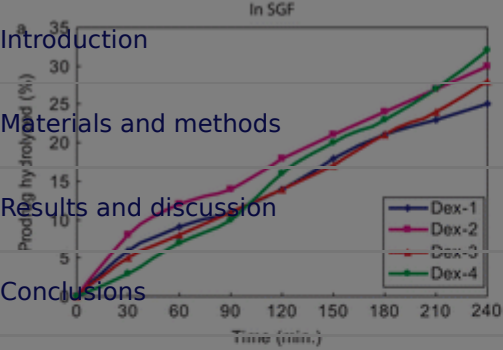
acacia a standard group, e sed in acrylic c midity of 45- 55%, in with standard rodent d ek before experi lines of the Com val of the Institutio acy,

Tirupati,

Anti-in

This study describes the synthesis of amide prodrugs of Dex as Dex-1 {S(+)-methyl-3-(1H-indol-3-yl)-2-(2-(4-isobutylphenyl)propanamido)propanoate}, Dex-2 {S(+)-methyl-2-(2-(4-isobutylphenyl)propanamido)-3-phenylpropanoate}, Dex-3 {S(+)-methyl-2-(2-(4-isobutylphenyl)propanamido)acetate} and Dex-4 {S(+)-methyl-3-(4-hydroxyphenyl)-2-(2-(4-isobutylphenyl)propanamido)propanoate} as portrayed in [Scheme 1](#). The prodrugs were subjected to solubility, physicochemical characterization, protein binding and hydrolytic studies. All the prodrugs were observed to be highly soluble in organic solvents, sparingly soluble in alkaline solution and insoluble in acidic solutions. The high solubility of the prodrugs in organic solvents indicates their lipophilic behaviour. The physicochemical properties of synthesized prodrugs were determined, as well as the IR, ^1H NMR, ^{13}C NMR and mass spectral analyses were carried out. The results of elemental analysis of synthesized prodrugs were in all cases within $\pm 0.4\%$ of theoretical value and were in confirmation of desired structure.

In vitro hydrolysis of Dex-1, Dex-2, Dex-3 and Dex-4 were carried out in SGF (pH 1.2), SIF (pH 7.4), 80% human plasma (pH 7.4) and rat faecal matter (pH 7.4) and their respective half-life ($t_{1/2}$) values in SIF were observed as 4.5, 4.8, 4.5 and 4.6 h, respectively. The comparative hydrolysis patterns of the synthesized prodrugs are depicted in [Figure 1](#). None of the prodrugs showed significant hydrolysis in SGF. The amount of prodrugs hydrolyzed in SIF was found as 85%, 82%, 85% and 85%, respectively, in 80% human plasma and rat faecal matter. The hydrolysis patterns of the synthesized prodrugs encouraged the use of these prodrugs followed by Dex-1, Dex-2, Dex-3 and Dex-4. The hydrolysis of prodrugs showed that Dex-1 and Dex-2 showed 85% hydrolysis. This increase in hydrolysis required a lower dose will be required. Figure 1.



Display full size

Pharmacological studies

Anti-inflammatory activity

Table 2 shows that after 6 h of administration of Dex, the percentage anti-inflammatory activity was found as 43.3%, whereas an improved value of 73.4, 77.3, 72.8 and 64.5% were observed for prodrugs Dex-1, Dex-2, Dex-3 and Dex-4, respectively. The maximum anti-inflammatory activity of prodrugs was observed at 6 h and remained practically constant upto 8 h. The anti-inflammatory activity of free Dex decreased with time, whereas that of its prodrugs increased with time due to its higher bioavailability

comparative study showed that ANOVA showed no significant difference in the comparative study of prodrugs and free Dex. The results of the study are summarized in Table 2.



Download

Analgesic

In this article

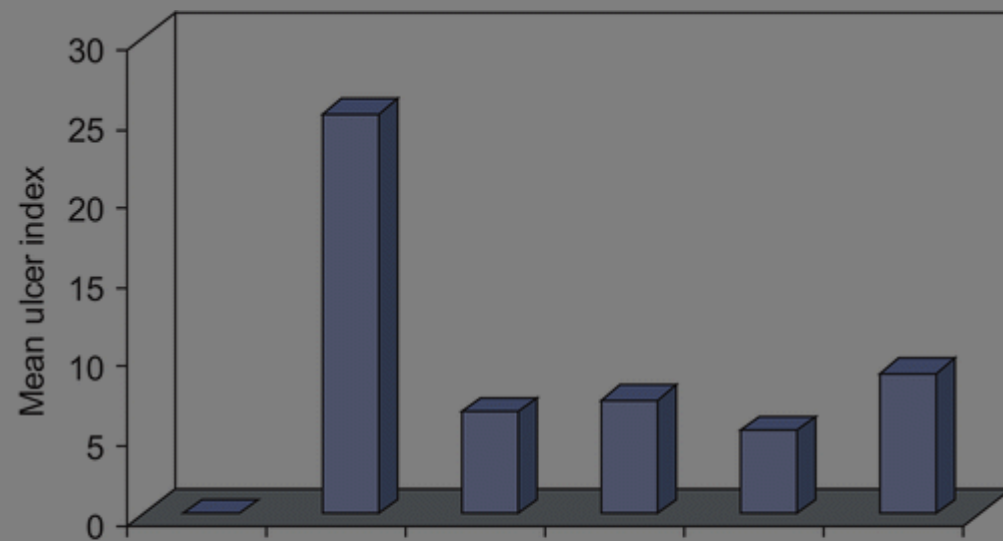


The percentage protection in mice brought about by administration of drug is shown in Table 2. After 4 h of the administration of Dex, the percentage anaesthesia was observed as 46.4%, whereas for Dex-1, Dex-2, Dex-3 and Dex-4, corresponding values of 61.4, 59.7, 63.6 and 52.7% were observed. The study revealed that analgesic activity of Dex decreased with time, while the prodrugs showed an improved analgesic activity.

Ulcerogenicity

Dex has produced a mean UI of 25.3 while that of its prodrugs resulted in producing very less values and is shown in Figure 2. The study revealed that the minimized side effect obtained in the prodrugs might be due to the inhibition of direct contact of carboxylic acid group of the drug to the gastric mucosa, which is mainly responsible for the damage.

Figure 2. Comparative ulcer index of dexibuprofen and its prodrugs.



Normal Control

Display full

History

The hist

tissues v

showed

revealed

The gastric

group rats

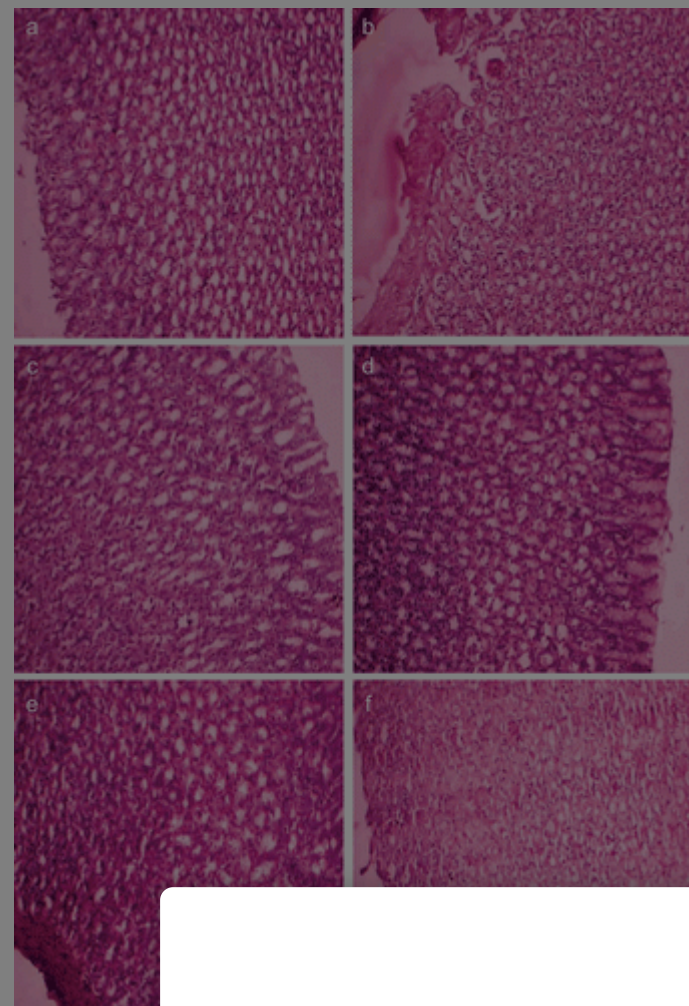
Dex group

the basal

In this article

stomach lumen. In this zone, the structures of the gland were destroyed. They had disintegrated from the basal lamina and fallen into the lumen. The nuclei of these cells became smaller and dense, and their cytoplasm was stained as dark eosinophilic bodies. Small hemorrhagic areas and patches of inflammatory cell infiltrations were present in the lumen of the glands and lamina propria. Normal histological findings were displayed for Dex-1, Dex-2, Dex-3 and Dex-4 revealing that the prodrugs are not producing any ulceration in the gastric region.

Figure 3. Histopathological studies of dexibuprofen and its prodrugs on rat stomach.



Display full



Con
In this st
the struc
studies o
organic

sized and
rolysis
bility in
t faecal

In this article



quantitatively in 80% human plasma, rat faecal matter and SIF, but were resistant to hydrolysis at SGF indicating that the prodrugs are resistant to acidic conditions. In vivo studies showed sustained release of Dex upto 8 h as also evident from the longer anti-inflammatory activity observed with prodrugs treated animals compared with Dex-treated animals. The prodrugs were found to be significantly less ulcerogenic and with more analgesia than parent drug. The histopathological study revealed limited ulcer formation in stomach by the prodrugs. These findings suggested that the prodrugs are better in action as compared with dexibuprofen and are advantageous in having less GI side effects without loss of desired anti-inflammatory and analgesic activity of the drug.

Acknowledgements

The authors thank M/s. Alkem Laboratories, Mumbai, India, for providing gift sample of dexibuprofen. The authors are grateful to Padmashree Dr. M. Mohan Babu, Chairman, Sree Vidyanikethan Educational Trust, Tirupati, India, for providing the necessary facilities to carry out this work.

Declaration of interest

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.



Rela

In sil



Linki

tion of

Refer

In this article



1. Kaehler ST, Phleps W, Hesse E. Dexibuprofen: pharmacology, therapeutic, use and safety. *inflammopharm* 2003;11:371-383.

Materials and methods

[PubMed](#) | [Google Scholar](#)

Results and discussion

2. Halan PK, Murumkar PR, Giridhar R, Yadav MR. Prodrug designing of NSAIDs. *Mini-Rev Med Chem* 2009;9:124-139.

Acknowledgements

[PubMed](#) | [Web of Science ®](#) | [Google Scholar](#)

References

3. Bajpai AK, Mishra A. Carboxymethyl cellulose (CMC) based semi-IPNs as carriers for controlled release of ciprofloxacin: an in-vitro dynamic study. *J Mater Sci Mater Med* 2008;19:2121-2130.

[PubMed](#) | [Web of Science ®](#) | [Google Scholar](#)

4. Mishra A, Ravichandran V, Jain PK, Dixit VK, Agrawal RK. Synthesis, characterization and pharmacological evaluation of amide prodrugs of ketorolac. *Europ J Med Chem* 2008;43:2464-2472.

[PubMed](#) | [Web of Science ®](#) | [Google Scholar](#)

5. Bhosle D, Bharambe S, Gairola N, Dhaneshwar SS. Mutual prodrug concept: fundamentals and applications. *Indian J Pharm Sci* 2006;68:286-294.

[Google Scholar](#)

6. Rasheed A, Kumar CKA. Novel approaches on prodrug based drug design. *Pharm Chem*

7. Roy S. (prodrug) through

8. Idle JR. cetic acids in the

9. Nielsen NW, Bundgaard H. Glycolamide esters as biolabile prodrugs of carboxylic acid agents: synthesis, stability, bioconversion, and physicochemical properties. *J Pharm Sci* 1988;77:285-298.

Results and discussion | PubMed | Web of Science ® | Google Scholar

Conclusions

10. Mohan R, Ramaa CS. Ester prodrugs of Flubiprofen: Synthesis, plasma hydrolysis and gastrointestinal toxicity. *Indian J Chem* 2007;46B:1164-1168.

References | Google Scholar

1. Dev S, Deepali VM, Kadam SS, Dhaneshwar SR. Synthesis and pharmacological evaluation of cyclodextrin conjugate prodrug of Mefenamic acid. *Indian J Pharm Sci* 2007;69: 69-72.

Google Scholar

2. Winter CA, Risely EA, Nuss GW. Carregeenan induced oedema in hind paw of the rat as assay for anti-inflammatory drugs. *Proc Soc Exp Biol Med* 1962;111:544-547.

PubMed | Web of Science ® | Google Scholar

3. Kulkarni SK. *Handbook of Experimental Pharmacology*, 3rd ed., Vallabh Prakash, New Delhi, 2001;128-130.

Google Scholar

4. Coili V, Putzolu S, Rossi V, Barcellona PS, Corradino C. The role of direct tissue contact in the ... rats. *Toxicol Appl P*

5. Yagmu ... ic oxide on ... :13-19.


Download

- Information for
 - Authors
 - R&D professionals
 - Editors
 - Librarians
 - Societies
 - Opportunities
 - Reprints and e-prints
 - Advertising solutions
 - Accelerated publication
 - Corporate access solutions

- Open access
 - Overview
 - Open journals
 - Open Select
 - Dove Medical Press
 - F1000Research
 - Help and information
 - Help and contact
 - Newsroom
 - All journals
 - Books

Keep up to date

Register to receive personalised research and resources by email

 Sign me up



Copyright © 2024 John Wiley & Francis Group
All rights reserved. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

Accessibility



Register
5 How

